

A FORENSICALLY RELEVANT UNSUPERVISED LEARNING APPROACH THAT ACCURATELY CLUSTERS SINGLE-CELL ELECTROPHEROGRAMS

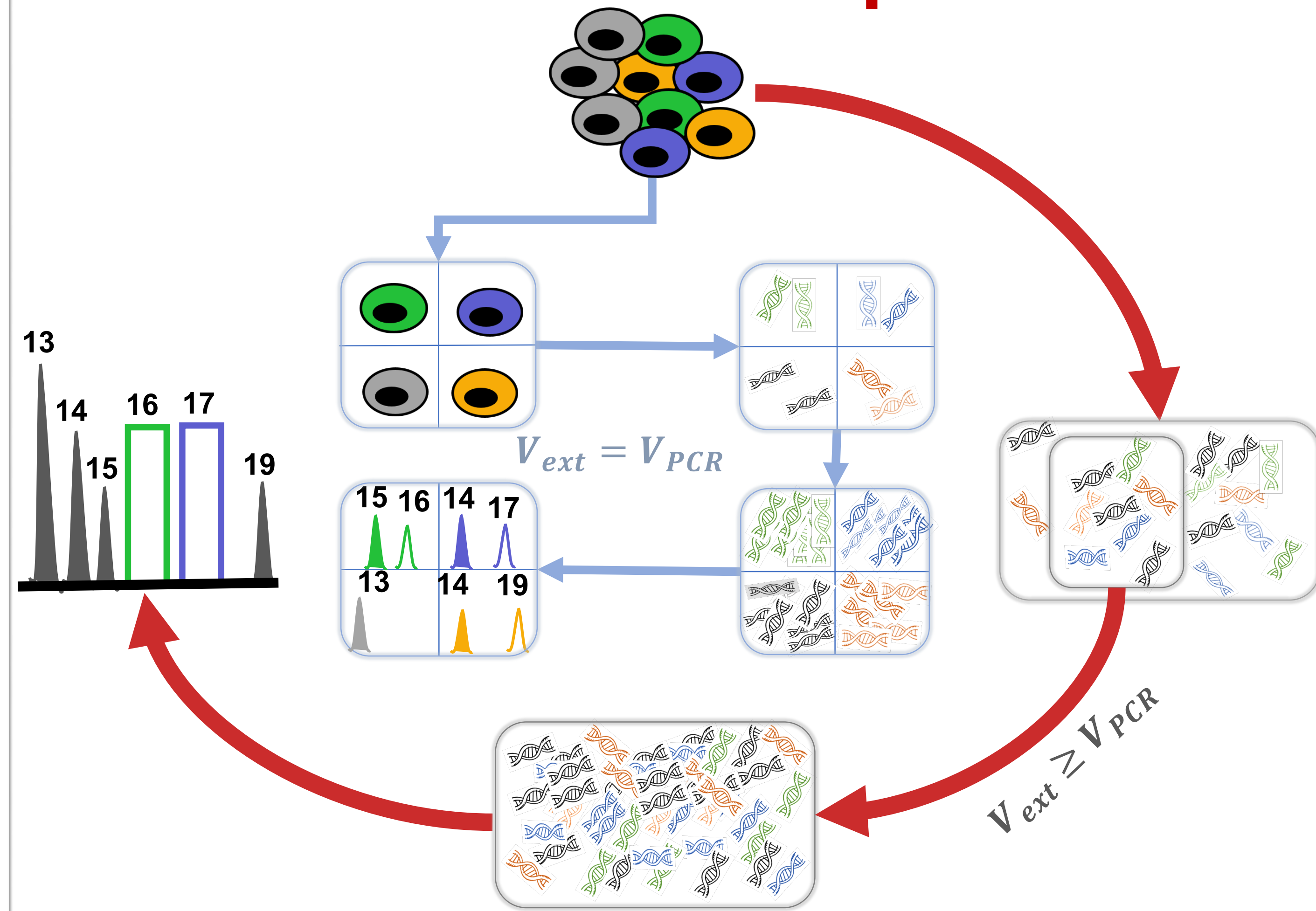
LFTDI

Nidhi Sheth ^{a,*}, Leah O'Donnell ^b, Madison Mulcahy ^a, Ken R. Duffy Ph.D. ^b, and Catherine M. Grgicak, Ph.D. ^a

^aRutgers University, Camden, US; ^bMaynooth University, Ireland



Mixture Interpretation of Bulk Samples vs. Single Cell Samples



Schematic of the single-cell (inner loop) and traditional bulk mixture (outer loop) pipelines.

In bulk mixture systems the cells are lysed, and the DNA is extracted into a final volume, V_{ext} . A portion of V_{ext} is transferred to a PCR tube, V_{PCR} , to which PCR reagents are added.

In contrast, single-cell systems isolate cells into distinct vessels prior to extraction, which is performed using direct-to-PCR reagents. The addition of PCR reagents to the entire V_{ext} follows, therein circumnavigating fractionation effects that lead to complete allele drop-out. Fragment analysis ensues, resulting in an EPG for each cell. Each EPG, therefore, is composed of only a single contributor's DNA.

Treatment Comparison

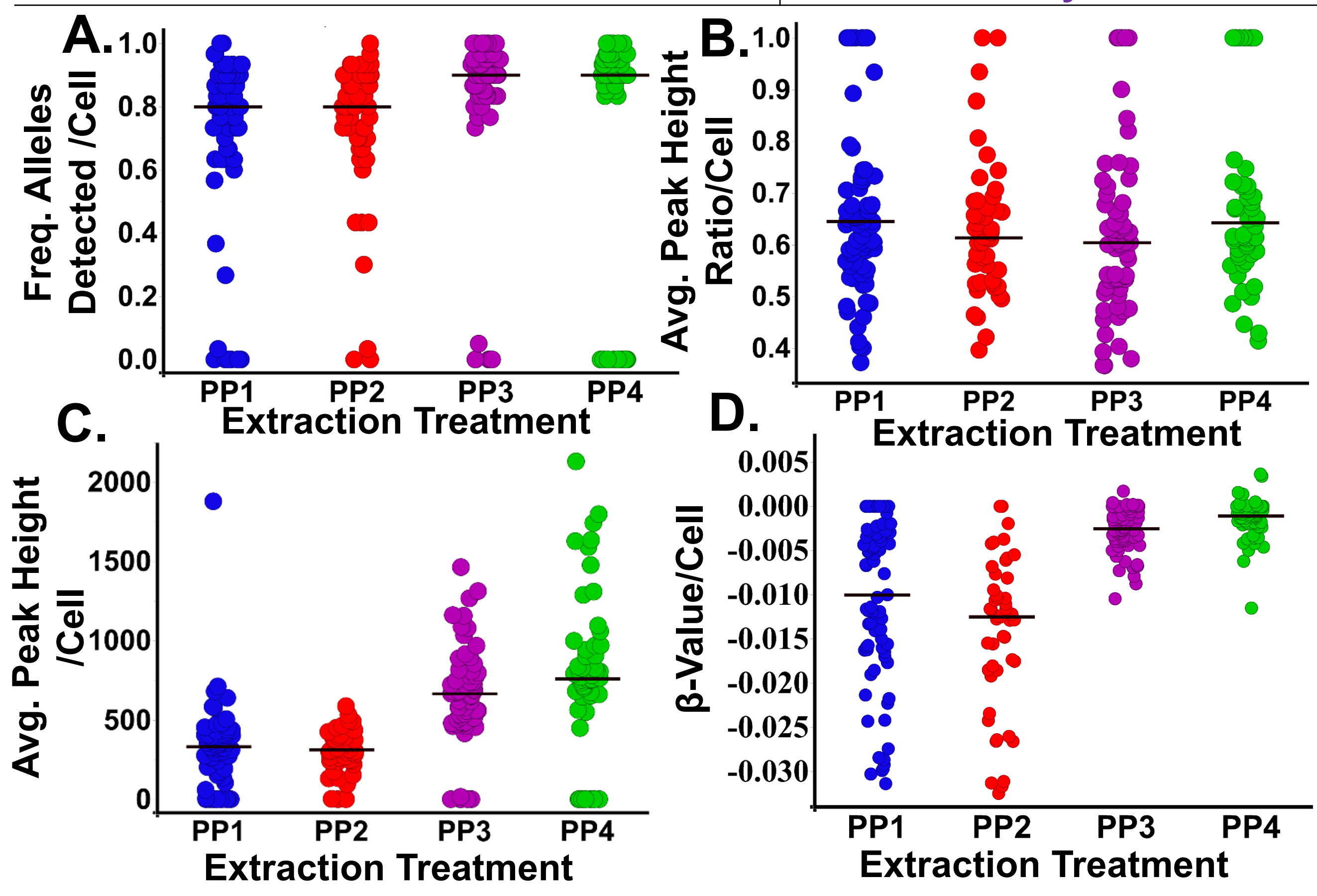
DEParray™ benchtop instrument combines microfluidics and microelectronics into an image-based system that isolates nucleated single-cells.

Extraction Treatment Variations

PP1	Higher Pro K and PBS (1X)
PP2	Lower Pro K and PBS (1X)
PP3	Lower Pro K and PBS (0.5X)
PP4	Lower Pro K and PBS (0.25X)

Experimental Conditions

Sample Set	241 Single-cell EPGs(scEPGs)
Amplification, Separation, Analysis	Global Filer™ (29 Cycles), 25 sec injection on 3500 Genetic analyzer, and Osiris (10.3.1) Analysis

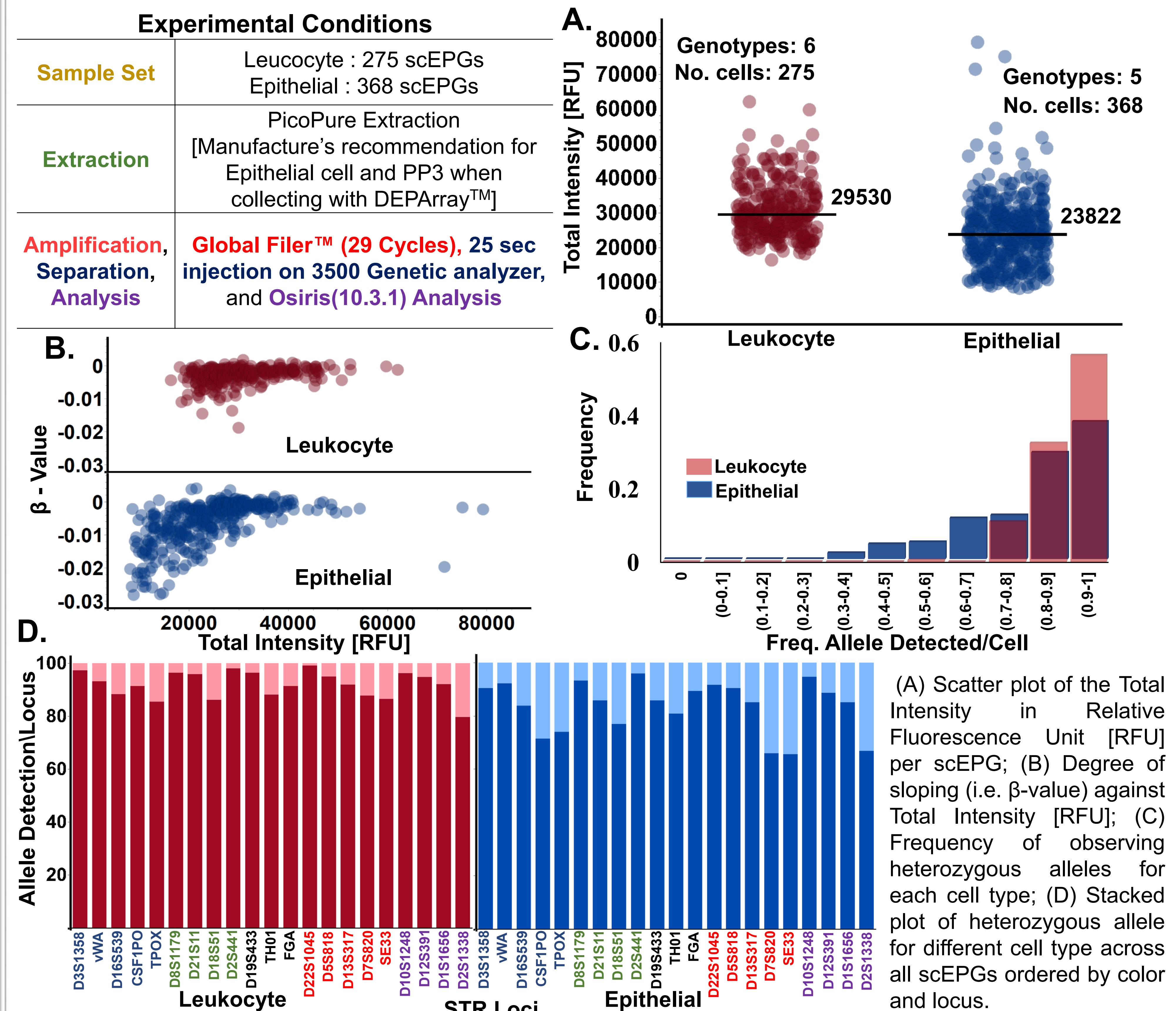


Extraction Treatment	(A)	(B)	(C)	(D)
PP1	0.8	0.645	333	-0.01
PP2	0.8	0.614	314	-0.012
PP3	0.9	0.604	667	-0.002
PP4	0.9	0.643	761	-0.001

(A) The frequency of detection for heterozygous alleles separated by extraction method; (B) Average peak height ratio per cell; (C) Average peak height per cell; (D) β -value – i.e., EPG sloping – per cell. The table shows the medians of each.

Single-Cell Exploratory Analysis

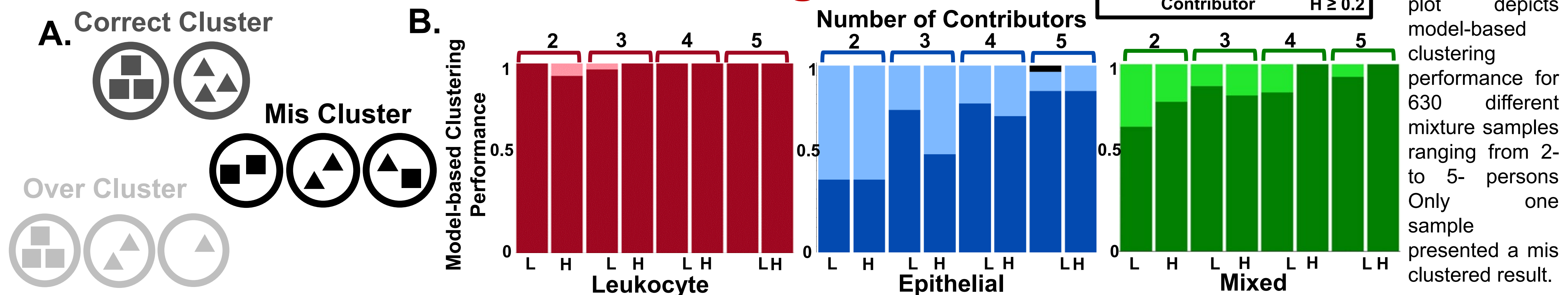
Exploratory analysis on single leukocytes and epithelial cells collected via two collection techniques: i) semi-automated DEParray™ NxT and ii) manual pico-pipetting technique.



(A) Scatter plot of the Total Intensity in Relative Fluorescence Unit [RFU] per scEPG; (B) Degree of sloping (i.e. β -value) against Total Intensity [RFU]; (C) Frequency of observing heterozygous alleles for each cell type; (D) Stacked plot of heterozygous allele for different cell type across all scEPGs ordered by color and locus.

We developed a method to cluster/group profiles by scEPG similarity, wherein the scEPGs of each group are then used to determine the Likelihood Ratio for a cluster of cells. Various methods were evaluated, with the most favorable being the model-based clustering as implemented in the R package *mclust* [1]. There are two types of clustering errors that might occur: i) overclustering, and ii) misclustering. Figure (A) is a schematic showing correct clustering, overclustering (known genotype is clustered in more than one group), and misclustering (where at least one group contains more than one genotype).

Model-Based Clustering



Conclusion

1. PicoPure™ DNA extraction treatment coupled with a lower concentration of PBS and Pro K led to higher peak heights, larger allele detection rates, and lower sloping values.
2. MBC, as implemented in *mclust*, showed promising results as it correctly clustered samples consisting of 2- to 5- person mixtures with only one exception.

Reference

1. L. Scrucca, M. Fop, T. B. Murphy, and A. E. Raftery, "Mclust 5: Clustering, classification and density estimation using Gaussian finite mixture models," *The R Journal*, vol. 8, no. 1, pp. 289-317, 2016.

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